



Anti-GFP MagBeads

Product description

Anti-GFP MagBeads are magnetic polymer beads. They are made of high quality murine monoclonal antibodies and carboxyl magnetic beads. They provide rapid magnetic responsiveness. Superparamagnetism, good dispersion, uniform particle size, very low non-specific binding and abundant binding sites. It can be used for GFP labeled protein immunoprecipitation (IP).

Components

Components No.	Name	20564ES76	20564ES03	20564ES08
20564	Anti-GFP MagBeads	500 μ L	1 mL	5 mL

Specifications

Ligand	Anti-GFP Antibody
MagBeads Concentration	10 mg/mL
Protein binding capacity	\geq 0.6 mg protein/mL
Storage buffer	ddH ₂ O

Storage

The products should be stably stored at 2~8°C for 1 year. Do not freeze!

Instructions

1. Sample preparation

Note: The sample may be a bacterial fermentation broth or a cell lysate. It should not contain insoluble particles, which can be filtered by 0.22 μ m filter membrane or centrifuged by 10,000 \times g for 15 min.

2. Preparation of Magnetic Beads

2.1 Resuspend the Magnetic Beads in the vial(tilt and rotate for 2 minutes or gently pipette for 10 times).Transfer 25-50 μ L of Anti-Flag MagBeads into a 1.5mL tube(Transfer amount maybe adjusted as required).

2.2 Add 500 μ L of IP Lysis/Wash Buffer to the beads and gently pipette to mix. Place the tube into a magnetic stand to collect the pipette to mix.Place the tube into a magnetic stand to collect the Beads against the side of the tube(Herein after referred to as Magnetic separation).Remove and discard the supernatant.

2.3 Repeat step 2.2 twice.

3. Immunoprecipitation

3.1 Remove the tubes from the magnetic separator and add your sample containing Flag-tagged protein to the pre-washed magnetic beads and incubate at room temperature for 2 h or overnight at 4°C with mixing.

3.2 Collect the beads with a magnetic stand, remove the unbound sample and collect for analysis.



3.3 Add 500 μ L of IP Lysis/Wash Buffer to the tube and gently mix. Collect the beads and discard the supernatant. Repeat wash twice.

4. Elution

4.1 Acidic Elution

4.1.1 Add 100 μ L of 0.1 M glycine, pH3.0.

4.1.2 Gently vortex to mix and incubate the sample at room temperature on a rotator for 5-10 minutes.

4.1.3 Magnetically separate the beads and save the supernatant containing the target antigen.

4.1.4 To neutralize the low pH, add 20 μ L of Neutralization Buffer (1 M Tris pH 8.5) for each 100 μ L of eluate. The final solution can be used as samples for denaturing SDS-PAGE.

4.2 Basic Elution

4.2.1 Add 100 μ L of 1xSDS-PAGE loading buffer to the tube.

4.2.2 Boil for 5 minutes on a dry bath.

4.2.3 Magnetically separate the beads and the supernatant was taken for SDS-PAGE detection..

Notes

1. For your safety and health, please wear lab coats and disposable gloves for operation.
2. For research use only.